

REVIEW PAPER

Tomasz Kubrak (D 1,2 (ABCDEFGH), Rafał Podgórski (D 1,2 (ABCDEFGH), Monika Stompor (D 1,2 (ABCDEFGH)

Natural and Synthetic Coumarins and their Pharmacological Activity

¹ Centre for Innovative Research in Medical and Natural Sciences, University of Rzeszów ² Department of Biochemistry, Faculty of Medicine, University of Rzeszów

ABSTRACT

Coumarins are a structurally diverse group of natural substances derives from plants that display a host of bioactivities. In this paper, we will introduce the reader to coumarins and their applications as medicinal substances. The great diversity in courmarin structure will be discussed along with their extensive use as pharmaceutical agents. Coumarins display a wide range of antimicrobial activity and applications of coumarins as antifungal and antiviral agents will be addressed. Other properties of coumarins such as their role in neuroprotection, anticancer, and as antioxidants will also be reviewed.

Keywords. coumarins, antimicrobial agents, neuroprotection, natural products in medicine

Introduction

Coumarins form an extensive group of natural substances known as secondary metabolites. They are found in over 150 different species of plants belonging to almost 30 different families. The families containing the highest content of coumarins are: *Rutaceae*, *Clusiaceae*, *Guttiferae*, *Caprifoliaceae*, *Oleaceae*, *Nyctaginaceae* and *Apiaceae*. Coumarin compounds accumulate in large quantities in fruits (such as citrus fruits), vegetables (eg. celery), roots, flowers and leaves. In smaller quantities they are isolated from bark and stems.

Coumarins, in addition to occurring in vascular plants, are also found in bacteria and fungi such as novobiocin and coumermicin which are known antibiotics synthesized by bacteria. In contrast, *Aspergillus flavus* is a source of aflatoxin, a highly carcinogenic substance with acoumarin ring in its structure.³

Structural diversity of coumarins

The structural diversity of natural coumarins is the basis for classifying them into four groups:

1. **coumarin derivatives**, e.g. simple **coumarin**, compounds formed by two rings: benzene and α-piron. Substituents are often hydroxyl, methoxy and aliphatic groups, at the C7, C6 and C3 positions of benzopyrone (Fig. 1).

Figure 1. Chemical structure of coumarin

Corresponding author: Tomasz Kubrak, e-mail: tkubrak@ur.edu.pl

Participation of co-authors: A – Author of the concept and objectives of paper; B – collection of data; C – implementation of research; D – elaborate, analysis and interpretation of data; E – statistical analysis; F – preparation of a manuscript; G – working out the literature; H – obtaining funds

Received: 12.02.2017 | Accepted: 06.06.2017

Publication date: June 2017

Kubrak T, Podgórski R, Stompor M. *Natural and Synthetic Coumarins and their Pharmacological Activity.* Eur J Clin Exp Med. 2017;15(2):169–175. doi: 10.15584/ejcem.2017.2.12

2. **isocoumarin derivatives**, formed by benzene rings and α-isopirone. They have substituents in positions C3, C6, C7 and C8 (Fig. 2). They are isolated mainly from fungi: *Artemisia, Aspergilus, Fusarium, Penicillinium, Stremtomyces* and the few plants belonging to families: *Compositae, Leguminoseae* and *Myriaceae*.⁴

Figure 2. Chemical structure of isocoumarin

3. furanocoumarin derivatives, (Fig. 3) formed by the coupling of the coumarin ring with the furan ring at the C6-C7 position (psoralen type, Fig. 3A) or in the C7-C8 position (angelicin type, Fig. 3B).

Figure 3. Chemical structure of furanocoumarin

Figure 3A. Psoralentype

Figure 3B. Angelicintype

4. **pyrancoumarin derivatives**, coumarine ring is condensed with pyran ring (Fig. 4). Ring condensation at the C6-C7 position is defined by the xanthyletin-type (Fig. 4A), or in position C7-C8 a seselin-type (Fig. 4B).

Figure 4. Chemical structure of pyrancoumarin

Figure 4A. Xanthyletin-type

Figure 4B. Seselin-type

Pharmacological Activity of Coumarins

Coumarins are a group of biologically active compounds. They are produced by living organisms (plants, fungi and bacteria) as secondary metabolites. Their activities are, among others, anti-inflammatory, antithrombotic, antimicrobial, antifungal, antiviral (including anti-HIV), anticonvulsant, antioxidant, and antitumor.¹

Anti-inflammatory activity of coumarins

Coumarins (1,2-benzopyrone) haveanti-inflammatory properties andhavebeen used to treat oedema, helping wound healing. This removes protein and oedemafluid from injured tissue by stimulating phagocytosis and proteolytic enzyme production.⁵ Esculetin exhibited anti-inflammatory activityin rat colitis.⁶ Also, esculetin inhibits the cyclooxygenase and lipoxygenase enzymes, which results in an anti-inflammatory effect.⁷

Anticoagulant activity of coumarins

Vitamin K is a co-catalyst for the carboxylation reaction of the glutamic acid residue with γ -carboxyglutamic acid. The carboxylation process affects the further normal activity of coagulation factors II, VII, IX and X. Warfarin interferes with the cycle of vitamin K metabolism, resulting in liver deposition of partially carboxylated and decarboxylated proteins. These proteins are characterized

by decreased procoagulant activity.⁸ Coumarins interfere with the carboxylation process of C and S protein, causing a procoagulant effect.¹

It has been shown that the warfarin - coumarin derivative, used as an oral anticoagulant, negatively affects the γ -carboxylation of glutamate residues of bone proteins. As a result of its action in pregnant mothers and those taking warfarin preparation, the fetal skeleton develops abnormally.

Warfarin has shown particularly promising results in the treatment of SCCL (Small Cell Carcinoma Lung) a tumour cell type that is characterised by a coagulation-associated pathway.⁹

Antimicrobial activity of coumarins

Most coumarins have very low antimicrobial activity, but compounds having long chain hydrocarbon substitutions such as ammoresinol and ostruthin have a high activity towards gram(+) bacteria and show antimicrobial activity on Bacillus megaterium, Micrococcus luteus, Micrococcus lysodeikticus and Staphylococcus auerus.

Anthogenol, a coumarin derivative isolated from green fruits *Aegle marmelos* (L.), exhibits antimicrobial activity against bacteria of the genus *Enterococcus*. Imperatorinshows high activity towards *Shigella dysenteriae*. ¹⁰

Pyranocoumarins such as grandivittin, agasyllin, aegelinol benzoate and osthole isolated from the root *Ferulagocampestris*(Besser) Grecescu(*Apiaceae*) show activity towards both gram(+) andgram(-)bacteria, for example on *Staphylococcus aureus*, *Salmonella typhi*, *Enterobacter cloacae*, *Enterobacter aerogenes* and *Helicobacter pylori*.¹¹

Coumarins are mainly isolated from higher plants, but some of them have been discovered in microorganisms. Examples include novobiocin, coumermycin, and chartreusin. Novobiocin, a secondary metabolite of Streptomyces niveusand Streptomyces spheroidesexhibit very high activity towards gram(+) organisms such as Corinebacterium diphtheria, Staphylococcus aureus, Streptomyces pneumoniae, Streptomyces pyogenesandgram(-) organisms such as Haemophillusinfluenzae, Neisseria meningitides and Pasteurella. Coumermycin, structural similar to novobiocin, exhibits almost 50 times more potency againstbacteria belonging to Escherichia coli strains and Staphylococcus aureusstrains. Alsochartreusin, isolated from Streptomyces chartreusis, shows activity towards gram(+) bacteria, but due to its toxicity, chartreusin has not been tried for treatment.1 Antituberculous activity against Mycobacterium tuberculosisare found inskopoletin, umbelliferone, phellodenol A, marmezin and xanthyletin.¹²

Antifungal activity of coumarins

The broad spectrum of antifungal activity is shown byosthol, a derivative of coumarin isolated from celery plants. This derivative demonstrates activity against *Rhizoctoniasolani* [Kühn], *Phytophthoracapsici* [Leonian], *Botry-*

tis cinerea [Pers.], Sclerotiniasclerotiorum [de Bary] and Fusarium graminearum [Patch].¹³ A number of coumarins have been tested for antifungal activity, and the three most effective ones are psoralen, imperatorin, and ostruthin.¹⁴

Antiviral activity of coumarins

Coumarin derivatives can also have a reverse transcriptase (RT) inhibitory effectand two isomers (+)-calanolide A and (-)-calanolide B isolated from *Calophyllumlanigerum* (*Clusiaceae*), belong to pyranocoumarin, inhibit RT activityandcompletely deactivate the replication process of HIV-1. ^{15,16} Others coumarin derivatives – inophyllum B i P obtained from an giant African snail *Achatinafulica* [Férussac] significantly inhibit RT activity in HIV-1 cell cultures. ¹⁷

Aminomethyltrimethyl psoralen (AMT) is usedas a photo-sterilizing agent and is added to blood products followed by exposure to UVA radiation. AMT has inactivated DNA and RNA viruses. ¹⁸ Sancho et al. showed that imperatorin also inhibits either vesicular stomatitis virus or gp-160-enveloped recombinant HIV-1 infection in several T-cell lines and in HeLa cells. ¹⁹

Antihypertensive activity of coumarins

Scopoletin, a coumarin isolated from the fruits of *Tet-rapleuratetraptera* (*Mimosaceae*), in laboratory animals shows a smooth muscle relaxant effect onblood vessels resulting in a drop in blood pressure. A similar action wasshown by dihydromammea; a coumarin isolated from the seed of the tree *Mammeaafricana* [L.] (*Guttiferae*).¹

Visnadine – pyranokumarin, an active ingredient extracted from the fruit of *Ammi visnaga*, exhibited peripheral and coronary vasodilator activities. It is used adjunctively to treat angina pectoris.²⁰

Antioxidant activity of coumarins

Coumarin antioxidant activity is manifested by the ability to inhibit reactive oxygen species(ROS) and to capture them. Studies in rats have shown that the inhibition of xanthine oxidase - the enzyme responsible for xanthine biosynthesis, is directly proportional to the amount of hydroxyl groups that are contained withinthe molecule.²¹

Coumarin compounds can directly affect the properties of antioxidant enzymes. Łuczajet al. have demonstrated the effect of coumarin on superoxide dismutase, catalase, and glutathione peroxidase activity in plasma, liver, kidney and brain of rats. ^{22,23} Following administration of esculetin and 7-hydroxycoumarin in mice, increased levels of vitamin E, vitamin C and glutathionewere found. ²⁴ It has been shown that fraxin in 0.5 mM concentration protects human umbilical vein endothelial cells (HUVEC) against oxidative stress caused by hydrogen peroxide. ²⁵

The antioxidative and cytopreparative character of fraxetin has been demonstrated. It effectively prevents

oxidative stress-induced apoptosis of neuroblastoma cells-which can be used in the treatment of Parkinson's disease and other neurodegenerative diseases.²⁶

Neuroprotective activity of coumarins

Alzheimer's disease (AD) is a degenerative and progressive neurological disorder. It is characterized by variable levels of cholinergic enzymes and the formation of senile plaques containing β -amyloid proteins in cerebral tissue. In patients with Alzheimer's disease is observed decreased or unchanging levels of acetylcholinesterase (AChE), level of second enzyme butyrylcholinesterase (BChE) increase. Therefore, the levels of AChE and BChE enzymes are considered to be crucial in the treatment of this disease. Orhanet al.have demonstrated significant inhibition of acetyl- and butyrylcholinesterase levels after application with bergapten, xanthotoxin, scopoletin, umbelliferone, and 4-methyloumbelliferone.

Recent computer techniques have allowed the design of an amine-substituted coumarin derivative. The synthesized compound 3-(4-{[benzyl(ethyl)amino]methyl} phenyl)-7-[4-(diethylamino)butoxy]-2H-chromen-2-one exhibits neuroprotective activity, expressed in AChE inhibition, and is a potential candidate for Alzheimer's disease treatment.²⁸

Ostholepresent among others in *Cnidiummonnieri* (L.) fruits is a commonly used substance in traditional Chinese medicine. Chen et al. investigated the effect of osthole on the demyelination process in the central nervous system of mice in an experimental model of multiple sclerosis.²⁹ The results showed that osthole delayed disease progression and could find use in the treatment of multiple sclerosis.²⁹

The use of coumarins in the treatment of skin diseases and of the hematopoietic system diseases

Therapeutic use has been found for two furanocoumarin derivatives 5-MOP (5-methoxypsoralen) as an N-acetyl-transferaseinhibitor and 8-MOP (8-methoxypsoralen)in phototherapy for psoriasis and vitiligo.

In treatment of the skin disordersvitiligo, psoriasis and atopic inflammation, bergapten has also been successful. 30,31 Human keratinocytes of the NCTC-2544 line were exposed to bergaptene and xanthotoxinand exposed to UVA light, which resulted in cell cycle inhibition in G1 phase and increase in cellular apoptosis level. 32

Very effective psoriasis treatment was achieved using xanthotoxin and the PUVA method which involves administering xanthotoxin gel directly onto the patient's skin and then irradiating with UVB light.^{33,34}

The Jurkat cell line (T-cell leukemia line) and normal lymphocytes were exposed to 8-MOP and then exposed to UVA light. There was a marked induction of apoptosis and a significant increase in caspases: 8 and 9 (initiator

caspases) and 3 and 7 (effector caspases).³⁵ This method, called photophoresis, which uses extracorporeal irradiation of blood cells previously exposed to 8-MOP, has been implicated in therapy forautoimmune diseases, such as T-cell lymphoma.³⁶ Photophoresis increases apoptosis in lymphocytes, causing them to die and induce the formation of postapoptotic vesicles with anti-inflammatory properties.³⁷ Another feature of xanthotoxin used in vitiligo treatment is the ability to induce skin repigment. Coumarin increases the intracellular concentration of calcium ions and affects the organization of actin fibers in the cytoskeleton of melanocytes, which in turn leads to their migration.³⁸

Anticancer activity of coumarins

In research on GLC₄ (small cell lung carcinoma) and COLO 320 (colorectal cancer) cell lines, it has been shown that the cytotoxity of coumarin is due to the presence of at least two phenolic groups at the 6,7- or 6,8-position in the ring of the molecule.³⁹ The proliferation of the 786-O and A-498 (kidney cancer) and DU145 and LNCaP (prostate cancer) cells line were inhibited by coumarin and its hydroxyl derivative, umbelliferone.^{40,41}

Several hydroxylated and methoxylatedcoumarin derivatives were tested for their relative cytotoxicity on four human HSC-2 tumor cell lines, HSC-3 (oral squamous cell carcinoma), A-375 (melanoma) and HL-60 (promyelocytic leukemia). It has been shown that the cytotoxicity of 6,7-dihydroxycoumarin towards HL-60 tumor cells can be further enhanced by substituting the -OH in 3 and/or position 4.⁴² Similar conclusions were made by Budzisz et al.byQSAR regression analysis of the relationship between biological activity and physicochemical properties of test compounds. The cytotoxic effect increases with increasing hydrophobic substituents in 2, 3 and 4 positions of the benzopyrene ring.⁴³

The cytotoxic activity of organometallic coumarin complexes (umbelliferone, mendiaxon, warfarin, coumachlor, and niffcoumar) towards P3HR1, K-562 and THP-1 leukemia cells lines were confirmed.⁴⁴Nitrocoumarin derivatives of 7-hydroxy-6-nitrocoumarin and 7-hydroxy-3,6,8-trinitrocoumarin exhibited cytotoxic activity against tumor cells of the melanocytic line (SK-MEL-31).⁴¹ On the other hand, 8-nitro-7-hydroxy-coumarin induced apoptosis of leukemic cell lines K562 and HL-60.⁴⁵

Coumarin derivatives exhibit specific cytotoxicity, which is closely related to the chemical structure of their molecules. Attemptshave been made to synthesize a coumarin-like compound with selective and targeted action on tumor cells. Extremely cytotoxic heterocyclic coumarin derivatives which have 1,2,4-triazole, 4,5-dicyanoimidazole or purine groups have been obtained. In addition, the 1,2,4-triazole-3-carboxamide derivative exhibited particular selectivity to HeLa human epithelial

cells (cervical cancer).⁴⁴ In contrast, the presence of the 2-amino-6-chloropurine group conditioned the cytostatic effects on HepG2 (hepatoma) and SW620 (colon) cells line, leading to mutations in the p53 gene.⁴⁶

Osthole stops proliferation of human breast cancer cells MCF-7 and MDA-MB231 by inhibiting metalloproteinases in the outer cell matrix, slowing down the migration and further invasion of tumor cells.^{47,48}

Grandivittin, agasyllin, aegelinol benzoate and felamidin, four natural coumarins isolated from *Ferulagocampestris* (*Apiaceae*), and several synthetic ester derivatives of aegelinol were tested against four tumor cell lines. Some of them were shown to be marginally cytotoxic against the A549 lung cancer cell line.⁴⁹ From *Canophyllumdispar* (*Clusiaceae*), eight 4-phenylfuranocoumarin derivatives were isolated that showed significant cytotoxicity to cervical cancer (KB).⁵⁰

Panno et al. stimulated bergapten on breast cancer cells MCF-7 (human adenocarcinoma cell line) and SKBR-3 (malignant breast cancer cell line). Bergapten, independently of photoactivation, caused cell cycle arrest in G0 / G1 phase, inserting breast cancer cells into the pathway of apoptosis, and counteracting the stimulating effect of IGF-I/E2 on MCF-7 cell line growth.⁵¹ Further study of the team, conducted on human breast cancer cells MCF-7, ZR-75 and SKBR-3, confirmed the antiproliferative and apoptotic effects of bergapten and its UV-activated derivative.⁵² Molecular studies of mammary gland cells have determined the function of the membrane estrogen receptor a (ERa). ERa is involved in the normal development of the mammary gland as well as in the tumor-resistant MCF-7 breast cancer resistant to tamoxifen. Stimulation with bergapten causes ERa to decrease with anti-tumor and mitogenic effects.⁵³ Recent studies show that bergapten induces the metabolic reprogramming of breast cancer cells MCF-7 and ZR75. Therapy with bergapten causes changes in metabolic pathways, inducing cell death.54

Xanthoxylethin isolated from *Erythrinvariegata* [L.], stimulated gastric cancer cells of the SGC-7901 line, induced apoptosis and cell cycle arrest. It was noted that this action was associated with DNA damage. The process of apoptosis in cells was caused by mitochondrial damage and the cell cycle was stopped in phase S.⁵⁵

In cancer therapy, a very important issues are angiogenesis, i.e. the formation of blood vessels within the tumor and metastasis. The 7-diethylaminocoumarin derivatives exhibited activity as angiogenesis inhibitors towards to tumor cells and were highly selective to normal HUVEC (human umbilical vein endothelial cells). ⁵⁶ In other studies, the synthetic brominated coumarin derivative showed cytotoxic and anti-proliferative effects on EAC (Ascitic Carcinoma) and DLA (lymphoma) carcinoma cell lines. Inhibition of blood vessel formation and stimulation of apoptosis has also been observed. ⁵⁷

It has been observed that combination therapy with dicumarol, coupled with a chemotherapeutic agent, can improve efficacy and reduce toxicity compared to coumarin alone. The use of the dicumarol with taxol complex has antiproliferative effects on the hedgehog larvae (*Strongylocentrotuspurpuratus*) [Stimpson]. The positive result was explained by the synergism of the cytostatic and dicumarol. The authors suggest that the future of the development of combined pharmacotherapy may be the basis of modern chemotherapy.⁵⁸

It has been found that coumarins eaten in human diet can positively affect the body. Observations indicate that even if present at low levels in apiaceous vegetables, imperatorin, trioxsalen and isopimpinellin may contribute significantly to CYP1A2 inhibition and potentially decreased procarcinogen activation.⁵⁹

Conculsion

Coumarins are a large group of biologically active compounds commonly used in natural medicine.

References

- Venugopala KN, Rashmi V, Odhav B. Review on Natural Coumarin Lead Compounds for Their Pharmacological Activity. Bio Med Res Int. 2013;963248:1-14.
- Kohlmünzer S. Farmakognozja. Podręcznik dla studentów farmacji. Wydawnictwo Lekarskie PZWL;2000.
- Borges F, Roleira F, Milhazes N, Santana L, Uriarte E. Simple coumarins and analogues in medicinal chemistry: occurrence, synthesis and biological activity. CurrMed Chem. 2005;12:887-916.
- Superchi S, Phi D, Salvadori P, et al. Synthesis and Toxicity to Mammalian Cells of the Carrot Dihydroisocoumarins. Chem Res Toxicol. 1993;6:46-9.
- Piller NB. A comparison of the effectiveness of some anti inflammatory drugs on thermal oedema. Brit J ExperPath. 1975;56:554–60.
- Witaicenis A, Seito LN, Di Stasi LC. Intestinal anti-inflammatory activity of esculetin and 4-methylesculetin in the trinitrobenzenesulphonic acid model of rat colitis. Chem-Biol Interact. 2010;186:211–8.
- Fylaktakidou KC, Hadjipavlou-Litina DJ, Litinas KE, Nicolaides DN. Natural and synthetic coumarin derivatives with anti-inflammatory/antioxidant activities. Curr Pharm Design. 2004;10:3813–33.
- 8. Hirsh J, Dalen JE, Anderson DR, et al. Oral anticoagulants: mechanism of action, clinical effectiveness, and optimal therapeutic range. Chest. 1998;119:8–21.
- Lacy A, O'Kennedy R. Studies on Coumarins and Coumarin-Related Compounds to Determine their Therapeutic Role in the Treatment of Cancer. Curr. Pharm. Design. 2004;10:3797-811.
- 10. Raja SB, Murali MR, Roopa K, Devaraj SN. Imperatorin a furocoumarin inhibits periplasmic Cu-Zn SOD of *Shigelladysenteriae*their by modulates its resistance towards

- phagocytosis during host pathogen interaction. Biomed Pharmacother. 2011;5:560-8.
- 11. Basile A, Sorbo S, Spadaro V, et al. Antimicrobial and Antioxidant Activities of Coumarins from the Roots of *Ferulago-campestris* (Apiaceae). Molecules 2009;14:939-52.
- 12. Chiang CC, Cheng MJ, Peng CF, Huang HY, Chen IS. A novel dimeric coumarin analog and antimycobacterial constituents from *Fatouapilosa*. ChemBiodivers. 2010;7:1728-36.
- Wang CM, Zhou W, Li CX, Chen H, Shi ZQ, Fan YJ. Efficacy of osthol, a potent coumarin compound, in controlling powdery mildew caused by *Sphaerothecafuliginea*. J Asian Nat Prod Res. 2009;11:783-91.
- Bourgaud F, Hehn A, Larbat R, et al. Biosynthesis of coumarins in plants: a major pathway still to be unravelled for cytochrome P450 enzymes. PhytochemRev. 2006;5:293– 308.
- 15. McKee TC, Fuller RW, Covington CD, et al. New pyranocoumarins isolated from *Calophyllumlanigerum* and *Calophyllumteysmannii*. J Nat Prod. 1996;59:754-8.
- 16. Spino C, Dodier M, Sotheeswaran S. Anti-HIV coumarins from calophyllum seed oil. Bioorg Med Chem Lett. 1998;8:3475–8.
- 17. Patil AD, Freyer AJ, Eggleston DS, et al. The inophyllums, novel inhibitors of HIV-1 reverse transcriptase isolated from the Malaysian tree, *Calophylluminophyllum* Linn. J Med Chem. 1993;36:4131–8.
- Margolis-Nunno H, Robinson R, Ben-Hur E, Chin S, Orme T, Horowitz B. Elimination of potential mutagenicity in platelet concentrates that are virally inactivated with psoralens and ultraviolet A light. Transfusion 1995;35:855-62.
- Sancho R, Marquez N, Gomez-Gonzalo M, et al. Imperatorin inhibits HIV-1 replication through an Sp1-dependent pathway. J Biol Chem. 2004;279:37349–59.
- Iranshahi M, Askari M, Sahebkar A, Hadjipavlou-Litina D. Evaluation of antioxidant, anti-inflammatory and lipoxygenase inhibitory activities of the prenylated coumarinumbelliprenin. DARU J Pharm Sci. 2009;17:99–103.
- 21. Lee BC, Lee SY, Lee HJ, et al. Anti-oxidative and photo-protective effects of coumarins isolated from *Fraxinuschinensis*. Arch Pharm Res. 2007;30:1293-301.
- 22. Łuczaj W, Jarocka-Karpowicz I, Bielawska K, Skrzydlewska E. Sweet grass protection against oxidative stress formation in the rat brain. Metab Brain Dis. 2015;30:183–90.
- Łuczaj W, Stankiewicz-Kranc A, Milewska E, Roszkowska-Jakimiec W, Skrzydlewska E. Effect of sweet grass extract against oxidative stress in rat liver and serum. Food Chem-Toxicol. 2012;50:135-40.
- Martin-Aragòn S, Benedi JM, Villar AM. Effects of the antioxidant (6,7-dihydroxycoumarin) esculetin on the glutathione system and lipid peroxidation in mice. Gerontology. 1998;44:21-5.
- Whang WK, Park HS, Ham I, et al. Natural compounds, fraxin and chemicals structurally related to fraxin protect cells from oxidative stress. ExpMol Med. 2005;37:436–46.

- Molina-Jiménez MF, Sánchez-Reus MI, Andres D, Cascales M, Benedi J. Neuroprotective effect of fraxetin and myricetin against rotenone-induced apoptosis in neuroblastoma cells. Brain Res. 2004;1009:9-16.
- Orhan I, Tosun F, Sener B. Coumarin, anthroquinone and stilbene derivatives with anticholinesterase activity. Z Naturforsch C 2008;63:366-70.
- Montanari S, Bartolini M, Neviani P, et al. Multitarget strategy to address Alzheimer's Disease: design, synthesis, biological evaluation, and computational studies of coumarin-based derivatives. Chem Med Chem. 2016;11:1296-308.
- 29. Chen X, Pi R, Zou Y, et al. Attenuation of experimental autoimmune encephalomyelitis in C57 BL/6 mice by osthole, a natural coumarin. Eur J Pharmacol. 2010;629:40–6.
- 30. da Silva VB, Kawano DF, Carvalho I, da Conceição EC, de Freitas O, da Silva CH. Psoralen and bergapten: in silico metabolism and toxicophoric analysis of drugs used to treat vitiligo. J Pharm Pharm Sci. 2009;12:378-87.
- 31. Lohr *C*, Raquet N, Schrenk D. Application of the concept of relative photomutagenic potencies to selected furocoumarins in V79 cells. Toxicol In Vitro 2010;24:558-66.
- Viola G, Fortunato E, Cecconet L, Del Giudice L, Dall'Acqua F, Basso G. Central role of mitochondria and p53 in PUVA-induced apoptosis in human keratinocytes cell line NCTC-2544. ToxicolApplPharmacol. 2008;227:84-96.
- Asawanonda P, Amornpinyokeit N, Nimnuan C. Topical 8-methoxypsoralen enhances the therapeutic results of targeted narrowband ultraviolet B phototherapy for plaquetype psoriasis. J EurAcadDermatolVenereol. 2008;22:50-5.
- 34. SeckinD, UstaI, Yazici Z, Senol A. Topical 8-methoxypsoralen increases the efficacy of narrowband ultraviolet B in psoriasis. PhotodermatolPhotoimmunolPhotomed. 2009;25:237-41.
- Miyazaki M, Yamazaki H, Takeuchi H, Kamataki T. Mechanisms of chemopreventive effects of 8-methoxypsoralen against 4-(methylnitrosoamino)-1-(3-pyridyl)-1-butanone- induced mouse lung adenomas. Carcinogenesis. 2005;11:1947-55.
- Bissaccia E, Vonderheid EC, Geskin L. Safety of a new, single, integrated, closed photopheresis system in patients with cutaneous T-cell lymphoma. Br J Dermatol. 2009;161:167-9.
- 37. Stadler K, Frey B, Munoz LE, et al. Photopheresis with UV-A light and 8-methoxypsoralen leads to cell death and to release of blebs with anti-inflammatory phenotype in activated and non-activated lymphocytes. BiochemBiophys Res Commun. 2009;386:71-6.
- 38. Zhang XQ, Zheng M, Mou KH, Feng J. Effects of 8-Methoxypsoralen on intracellular Ca(2+)i and cytoskeleton actin organization in human melanocytes in vitro. J Zhejiang Univ Med Sci. 2009;38:348-51.
- Kolodziej H, Kayser O, Woerdenbag HJ, van Uden W, Pras N. Structure-cytotoxicity relationships of a series of natural and semi-synthetic simple coumarins as assessed in two human tumour cell lines. Z NaturforschC. 1997;52:240-4.

- Myers RB, Parker M, Grizzle WE. The effects of coumarin and suramin on the growth of malignant renal and prostatic cell lines. J Cancer Res ClinOncol. 1994;120:S11-3.
- Bielawska K, Malinowska M, Cyuńczyk M. Wpływ kumaryn na organizm człowieka. BromatChemToksykol. 2014;2: 213–21.
- 42. Kawase M, Sakagami H, Hashimoto K, Tani S, Hauer H, Chatterjee SS. Structure-cytotoxic activity relationships of simple hydroxylated coumarins. Anticancer Res. 2003;23:3243-6.
- 43. Budzisz E, Brzezinska E, Krajewska U, Rozalski M. Cytotoxic effects, alkylating properties and molecular modelling of coumarin derivatives and their phosphonic analogues. Eur J Med Chem. 2003;38:597-603.
- 44. Manolov I, Kostova I, Netzeva T, Konstantinov S, Karaivanova M. Cytotoxic activity of cerium complexes with coumarin derivatives. Molecular modeling of the ligands. Arch Pharm. 2000;333:93-8.
- 45. Egan D, James P, Cooke D, O'Kennedy R. Studies on the cytostatic and cytotoxic effects and mode of action of 8-nitro-7-hydroxycoumarin. Cancer Lett. 1997;118:201-11.
- 46. Benci K, Mandić L, Suhina T, et al. Novel coumarin derivatives containing 1,2,4-triazole, 4,5-dicyanoimidazole and purine moieties: synthesis and evaluation of their cytostatic activity. Molecules 2012;17:11010-25.
- 47. Yang D, Gu T, Wang T, Tang Q, Ma C. Effects of osthole on migration and invasion in breast cancer cells. BiosciBiotechBiochem.2010;74:1430-4.
- Zhang ZR, Leung WN, Cheung HY, Chan CW. Osthole: A review on its bioactivities, pharmacological properties, and potential as alternative medicine. Evid Based Complement Alternat Med. 2015;2015:919616.
- 49. Rosselli S, Maggio AM, Faraone N, et al. The cytotoxic properties of natural coumarins isolated from roots of *Ferulago-campestris* (Apiaceae) and of synthetic ester derivatives of aegelinol. Nat Prod Commun. 2009;4:1701-6.

- 50. Guilet D, Helesbeux JJ, Seraphin D, Sevenet T, Richomme P, Bruneton J. Novel cytotoxic 4-phenylfuranocoumarins from *Calophyllumdispar*. J Nat Prod. 2001;64:563-68.
- 51. Panno ML, Giordano F, Palma MG, et al. Evidence that bergapten, independently of its photoactivation, enhances p53 gene expression and induces apoptosis in human breast cancer cells. Curr Cancer Drug Targets 2009;9:469-81.
- 52. Panno ML, Giordano F, Mastroianni F, et al. Breast cancer cell survival signal is affected by bergapten combined with an ultraviolet irradiation. FEBS Lett. 2010;584: 2321-6.
- 53. Panno ML, Giordano F, Rizza P, et al. Bergapten induces proteasome-dependent degradation of ER in breast cancer cells: Involvement of SMAD4 in the ubiquitination process. Breast Cancer Res Treat. 2012;136:443–55.
- 54. Santoro M, Guido C, De Amicis F, et al. Bergapten induces metabolic reprogramming in breast cancer cells. Oncol Rep. 2016;35:568-76.
- Rasul A, Khan M, Yu B, Ma T, Yang H. Xanthoxyletin, a coumarin induces S phase arrest and apoptosis in human gastric adenocarcinoma SGC-7901 cells. Asian Pac J Cancer Prev. 2011;12:1219-23.
- 56. Lee S, Sivakumar K, Shin WS, Xie F, Wang Q. Synthesis and anti-angiogenesis activity of coumarin derivatives. Bioorg Med Chem Lett. 2006;16:4596-9.
- 57. Vijay Avin BR, Thirusangu P, Lakshmi Ranganatha V, Firdouse A, Prabhakar BT, Khanum SA. Synthesis and tumor inhibitory activity of novel coumarin analogs targeting angiogenesis and apoptosis. Eur J Med Chem. 2014;75: 211-21.
- 58. Madari H, Panda D, Wilson L, Jacobs RS. Dicoumarol: a unique microtubule stabilizing natural product that is synergistic with Taxol. Cancer Res. 2003;63:1214-20.
- 59. Kang AY, Young LR, Dingfelder C, Peterson S. Effects of furanocoumarins from apiaceous vegetables on the catalytic activity of recombinant human cytochrome P-450 1A2. Protein J. 2011;30:447-56.